

hMLH1/hMSH2

hMLH1/hMSH2 Protein Expression in Sporadic Colorectal Carcinoma and Its Clinicopathological Significance

Jae-Hee Kang, MD, Kil-Yeon Lee, MD, Kee-Hyu Lee, MD, Choong Yoon, MD, Soo-Mjung Ch, MD, Joo Hee Lee, MD.¹

Departments of Surgery and ¹Pathology, Kyung-Hee Univer Hspital, Seoul, Korea

Purpose : DNA replication errors (RERs) in repeated nu tide sequences (microsatellite instability) is cause tive mismatch repair (MMR) genes. Ninety percent colorectal carcinomas in hereditary nonpolyposis co cancer (HNPCC) patients and 10-15% of sporadic col rectal cancers show microsatellite instability. In t of colorectal cancers with microsatellite instabili fective MMR gene is hMLH1 or hMSH2. The author exam ined immunohistochemical expression of hMLH1 an hMSH2 in 75 cases of colorectal carcinomas exclu HNPCC, based on Amsterdam criteria for investigating copathological characteristics and prognosis in hMLH1 negative cases.

Methods : Formalin fixed, paraffin blocks obtaine tumors of 75 cases of colorectal cancers were staine two monoclonal antibodies (hMLH1 and hMSH2). The cor relation between hMLH1/hMSH2 negativity, and cli pathological feature and prognosis were statistica

Results : Twelve cases (16.0%) showed hMLH1/hMSH2 negativity. Negative expression of hMLH1/hMSH2 was sociated with early onset (under age 50), proximal l multiplicity, mucinous histologic type and poor d tiation. There was a significant survival advantage with hMLH1/hMSH2 negative colorectal carcinoma.

Conclusions : This study shows that hMLH1/hMSH2 neg tive colorectal carcinomas have the same clinicopath characteristics of colorectal carcinomas with micr instability. The immunohistochemical test for hMLH1/ protein can be a simple screening method routinely cable. The result of this test is available for e guidelines for managenent, and an independent progn factor for sporadic colorectal cancers. JKSCP 2001; 46

Key Words : Sporadic colorectal cancer, hMLH1, hMSH2, Mcrosatellite instability, Immunohistochemistry

1988 Vogelstein¹

가 (adenoma-carcinoma sequence)

(Hereditary Non-polyposis Colorectal Cancer, HNPCC)

deoxyribonucleic acid (DNA) (mismatch repair gene, MMR)

가 (replication errors, RER)가

DNA (microsatellite) mutation 가 가 (micro-

satellite instability, MSI)^{3,4} (Hereditary nonpolyposis col-orectal cancer, HNPCC)

90% MSI (germline mutation)가⁵⁻⁷

15% MSI^{8,9}

1 (: 130-702)
Tel: 02-958-8266, Fax: 02-966-9366
E-mail: keehlee@chollian.net

2)

가

^{3,10,11}

(1)

:

가

hMLH1/hMSH2

(poly-

가

merase chain reaction, PCR)
DNA

MSI test

가

(2)

:

가 가

hMLH1 monoclonal antibody (clone G 168-15, PharMingen, SanDiego, California, USA)

hMLH1 hMSH2

hMSH2 monoclonal antibody (clone G 219-1129, PharMingen, SanDiego, California, USA) 가

가

Avidin-Biotin-Peroxidase

가

가

4 μm

poly-L-lysine

MSI test

가

, PCR

coated glass slide

xylene

^{12,13}

. Endoge-

nous peroxidase

3% hy-

hMSH2

hMLH1/

drogen peroxidase

15

5

hMLH1/hMSH2

(antigen retrieval)

citrate buffer (pH

6.0) microwave 15

Tris

, hMLH1/hMSH2

buffer (pH 7.6) 가

hMLH1/hMSH2

가

hMLH1

hMSH2

1 : 50

37°C

가

가

가

1 biotinated antirabbit, anti-
mouse immunoglobulin

15

streptavidine

15

1)

. Diaminobenzidine

(chromagen)

1994

1

1995

6

hematoxylin

가

5

가

75

가

가

Amsterdam criteria¹⁴

가

. TNM

AJCC (American Joint Committee

on Cancer Criteria)

3) 56 (74.7%)
64 (85.3%),
11 (14.7%), 4 (5.3%),
hMLH1/hMSH2 7 (9.4%)
Pearson Chi-square test 가 5
Fisher's exact test P 0.05 33 (44.0%), 42 (56.0%)
69 (92%), 6
(8.0%)
5 hMLH1/hMSH2 25 (33.3%), 45 (60.0%),
Kaplan-Meier 5 (6.7%) 1 10
log-rank test (13.3%), 2 26 (34.7%), 3 35 (46.7%), 4
P 0.05 (5.3%) 가 4

2) hMLH1/hMSH2
hMLH1
가 9 (12%), hMSH2 가 4
(5.3%) hMLH1 hMSH2 가
33 82 , 58.3 가 1
47 (62.7%), 28 (37.3%) 89 hMLH1 hMSH2 가
19 (25.3%), 12 (16.0%)

Table 1. Comparison of clinicopathological features of hMLH1/hMSH2 negative and positive colorectal carcinoma

Features	Variables	Total number of cases	Number of hMLH1/hMSH2 negative cases (%) (n=12)	Number of hMLH1/hMSH2 positive cases (%) (n=63)	P value
Age	< 50 yr	19	7 (58.3)	12 (19.0)	0.004
	> 50 yr	56	5 (41.7)	51 (81.0)	
Sex	M	47	9 (75.0)	38 (60.3)	*N.S.
	F	28	3 (25.0)	25 (39.7)	
Location of tumor	Proximal	19	7 (58.3)	12 (19.0)	0.004
	Distal	56	5 (41.7)	51 (81.0)	
Multiplicity	Single	64	8 (66.7)	56 (88.9)	0.046
	Multiple	11	4 (33.3)	7 (11.1)	
Gross type	Exophytic	33	4 (33.3)	29 (46.0)	*N.S.
	Nonexophytic	42	8 (66.7)	34 (54.0)	
Histologic type	Non-mucinous	69	8 (66.7)	61 (96.8)	0.005
	Mucinous	6	4 (33.3)	2 (3.2)	
Diffrentiation	Well	25	3 (25.0)	22 (34.9)	0.021
	Moderate	45	6 (50.0)	39 (61.9)	
	Poor	5	3 (25.0)	2 (3.2)	
TNM stage	I	10	1 (8.3)	9 (14.3)	*N.S.
	II	26	6 (50.0)	20 (31.7)	
	III	35	5 (41.7)	30 (47.6)	
	IV	4	0 (0.0)	4 (6.3)	

*N.S. = not significant.

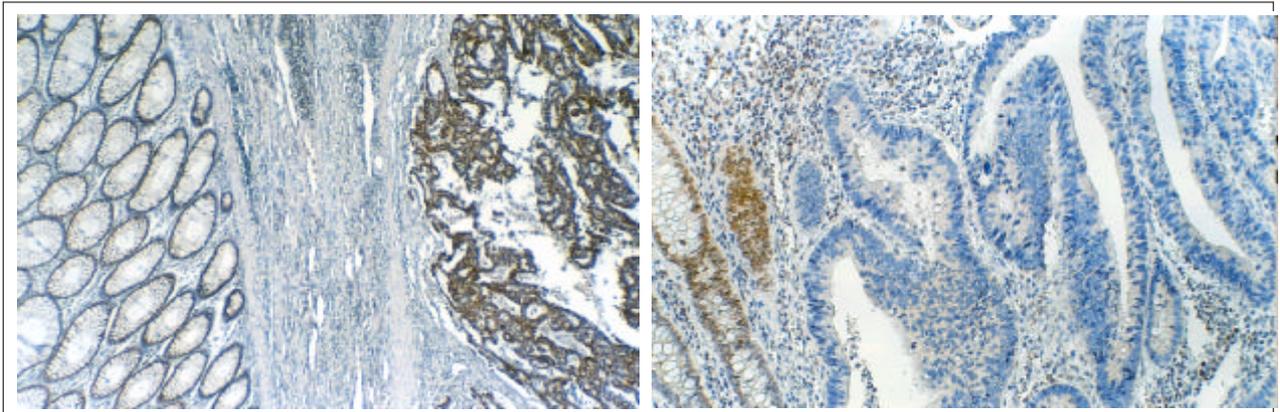


Fig. 3. Immunohistochemical staining of *hMLH1* and *hMSH2*. A. Diffuse, uniform nuclear staining of normal control mucosa and adjacent tumor cells with *hMLH1* ($\times 40$). B. Positive staining of normal control mucosa and lymphocytes with *hMLH1*; absence of staining in adjacent tumor nuclei is clearly evident ($\times 100$).

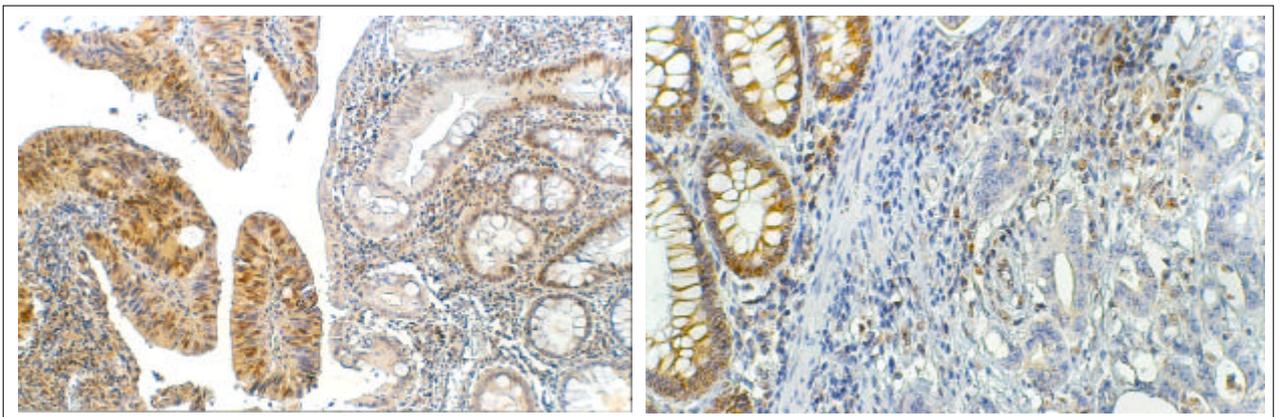


Fig. 4. A. Positive staining of normal control mucosa and adjacent tumor cells with *hMSH2* ($\times 100$). B. Loss of *hMSH2* expression in tumor cell. Normal mucosa, lymphocytes, and intervening stromal cells are positive ($\times 200$).

17

MSI

20% 10

15% 8,18 *hMLH1* *hMSH2*

가 가

Thibodeau¹⁸ low frequency MSI ()가 *hMLH1* *hMSH2*

hMLH1 *hMSH2* 가

3,10,11

human *mutL* homologue 1 (*hMLH1*), human *mutS* homologue 2 (*hMSH2*), *hMSH3*, *hMSH6*, *hMPS1*, *hMPS2* 가 가

hMSH2

가 가 40 60%

hMLH1

30% 5,16

MSI

high frequency MSI (MSI-H)

transforming growth factor (TGF-) (apoptosis) BAX

MSI 75% 100% PCR

12,13

test 가 MSI

가

hMLH1 hMSH2

sterdam criteria

hMSH2

가

(Bethesda guideline,²³ Japanese criteria²⁴)

가

hMLH1/hMSH2

ER/PR

가 가

가 Amsterdam criteria

가

hMLH1 /hMSH2

REFERENCES

1. Vogelstein B, Fearon ER, Hamilton SR, Kern SE, Preisinger AC, Lezppert M. Genetic alterations during colorectal tumor development. *N Engl J Med* 1988;319:525-32.
2. Kinzler KW, Vogelstein B. Lessons from hereditary colorectal cancer. *Cell* 1996;87:159-70.
3. Aaltonen LA, Peltomaki P, Leach FS, Sistonen P, Pylkkanen L, Mecklin JP, et al. Clues to the pathogenesis of familial colorectal cancer. *Science* 1993;260:812-6.
4. Boland CR, Thibodeau SN, Hamilton SR, Sidransky D, Eshleman JR, Burt RW, et al. A national cancer institute workshop on microsatellite instability for cancer detection and familial predisposition: development of international criteria for the determination of microsatellite instability in colorectal cancer. *Cancer Res* 1998;58:5248-57.
5. Liu B, Parsons R, Papadopoulos N, Nicolaides NC, Lynch HT, Watson P, et al. Analysis of mismatch repair genes in hereditary nonpolyposis colorectal cancer patients. *Nat Med* 1996;2:169-74.
6. Bronner CE, Baker SM, Morrison PT, Warren G, Smith LG, Lescoe MK, et al. Mutation in the DNA mismatch repair gene homologue hMLH1 is associated with hereditary nonpolyposis colorectal cancer. *Nature* 1994;368:258-61.
7. Fishel R, Lescoe MK, Rao MR, Copeland NG, Jenkins NA, Garber J, et al. The human mutator gene homologue MSH2 and its association with hereditary nonpolyposis colorectal cancer. *Cell* 1993;75:1027-38.
8. Lothe RA, Peltomaki P, Meling GI, Altonen LA, Nystrom-Lahti M, Pylkkanen L, et al. Genomic instability in colorectal cancer: relationship to clinicopathological variables and family history. *Cancer Res* 1993;53:5849-52.
9. Marra G, Boland CR. Hereditary nonpolyposis colorectal cancer: the syndrome, the genes and historical perspectives. *J Natl Cancer Inst* 1995;87:1114-25.
10. Kim H, Jen J, Vogelstein B, Hamilton SR. Clinical and pathological characteristics of sporadic colorectal carcinomas with DNA replication errors in microsatellite sequences. *Am J Pathol* 1994;145:146-56.
11. Horii A, Han HJ, Shimada M, Yanagisawa A, Kato Y, Ohta H, et al. Frequent replication errors at microsatellite loci in tumors of patients with multiple primary cancers. *Cancer Res* 1994;54:3373-5.
12. Victoria AM, Lisa M, Robert G, Hyeja K, Kelvin S, Anna M, et al. Immunohistochemistry for hMLH1 and hMSH2: a practical test for DNA mismatch repair-deficient tumors. *Am J Surg Pathol* 1999;23:1248-55.
13. Cawkwell L, Gray S, Murgatoryd H, Sutherland F, Haine L, Longfellow M, et al. Choice of management strategy for colorectal cancer based on a diagnostic immunohistochemical test for defective mismatch repair. *Gut* 1999;45:409-15.
14. Vasen HFA, Mecklin JP, Khan PM, Lynch HT. The international collaborative group on hereditary nonpolyposis colorectal cancer. *Dis Colon Rectum* 1991;34:424-5.
15. Jass JR, Do KA, Simms LA, Iino H, Wynter C, Pillay SP, et al. Morphology of sporadic colorectal cancer with DNA replication errors. *Gut* 1998;42:673-9.
16. Miyaki M, Konishi M, Muraoka M, Kikuchi-Yanoshita R, Tanaka K, Iwama T, et al. Germline mutations of hMSH2 and hMLH1 genes in Japanese families with

hereditary nonpolyposis colorectal cancer (HNPCC): usefulness of DNA analysis for screening and diagnosis of HNPCC patients. *J Mol Med* 1995;73:515-20.

(DNA sequencing)

가

17. Jonathan P, Terdiman MD, Peggy G, Conrad MS, Marvin HS. Genetic testing in hereditary colorectal cancer: indication and procedure. *Am J Gastroenterol* 1999;94:2344-56.

가

77%

18. Bubb VJ, Curtis LJ, Cunningham C, Dunlop MG, Carothers AD, Morris RG, et al. Microsatellite instability and the role of hMSH2 in sporadic colorectal cancer. *Oncogene* 1996;12:2641-9.

(MSI)

가 가

95%

MSI

19. Shigehiko M, Fumio K, Kazutomo T, Tomomi O, Shingo S, Kazuhisa S, et al. The significance of microsatellite instability in predicting the development of metachronous multiple colorectal carcinomas in patients with nonfamilial colorectal carcinoma. *Cancer* 1999;85:1917-24.

가

3%,

13 24%,

가

35%

MSI

MSI

20. Shingo S, Fumio K, Tomomi O, Hiroshi K, Kyotaro K, Michiko M, et al. Clinicopathologic and genetic features of nonfamilial colorectal carcinomas with DNA replication errors. *Cancer* 1998;82:279-85.

50%

MSH2

MLH1

가

6% MSI

가

MSI

가

MSI

21. Branch P, Hamson R, Karran P. DNA mismatch binding defects, DNA damage tolerance, and mutator phenotypes in human colorectal carcinoma cell lines. *Cancer Res* 1995;55:2304-9.

22. Boland CR. Roles of the DNA mismatch repair genes in colorectal tumorigenesis. *Int J Cancer* 1996;69:47-9.

TGF RII

APC, p53,

K-ras

LOH

23. Rodriguez-Bigas MA, Boland CR, Hamilton SR, Henson DE, Jass JR, Khan PM, et al. A national cancer institute workshop on hereditary nonpolyposis colorectal cancer syndrome: meeting highlights and Bethesda guidelines. *J Natl Cancer Inst* 1997;89:1758-62.

MSI가

MSI

9 17%

24. Kenichi K, Hideki M, Mariko E. Two types of sporadic multiple colorectal cancers with and without HNPCC-like genetic instability. *Hepatogastroentero* 1999;46:3115-3120.

MSI

가

DNA alkylation

가

G2/M check point

가

가

가

가

MSI

가 가

MSI

가

가

MSI

MSI

가

50%

가

MSI

hMLH1

hMSH2

가

가

가

MSI

MSI

MSI

MSI

MSI

hMLH1, hMSH2, hPMS1, hPMS2

hMLH1, hMSH2

hMLH1 hMSH2

가

MSI

MSI

